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Oxidative Stress and Cardiovascular Risk in Type 1 Diabetes Mellitus: Insights From the DCCT/EDIC Study

W.H. Wilson Tang, MD; Paula McGee, MS; John M. Lachin, ScD; Daniel Y. Li, BS; Byron Hoogwerf, MD; Stanley L. Hazen, MD, PhD; DCCT/EDIC Research Group*

Background—Hyperglycemia leading to increased oxidative stress is implicated in the increased risk for the development of macrovascular and microvascular complications in patients with type 1 diabetes mellitus.

Methods and Results—A random subcohort of 349 participants was selected from the DCCT/EDIC (Diabetes Control and Complications Trial/Epidemiology of Diabetes Interventions and Complications) cohort. This included 320 controls and 29 cardiovascular disease cases that were augmented with 98 additional known cases to yield a case cohort of 447 participants (320 controls, 127 cases). Biosamples from DCCT baseline, year 1, and closeout of DCCT, and 1 to 2 years post-DCCT (EDIC years 1 and 2) were measured for markers of oxidative stress, including plasma myeloperoxidase, paraoxonase activity, urinary F₂ iso prostaglandins, and its metabolite, 2,3 dinor-8 iso prostaglandin F₂α. Following adjustment for glycated hemoglobin and weighting the observations inversely proportional to the sampling selection probabilities, higher paraoxonase activity, reflective of antioxidant activity, and 2,3 dinor-8 iso prostaglandin F₂α, an oxidative marker, were significantly associated with lower risk of cardiovascular disease (−4.5% risk for 10% higher paraoxonase, \( P<0.003 \); −5.3% risk for 10% higher 2,3 dinor-8 iso prostaglandin F₂α, \( P=0.0092 \)). In contrast, the oxidative markers myeloperoxidase and F₂ iso prostaglandines were not significantly associated with cardiovascular disease after adjustment for glycated hemoglobin. There were no significant differences between DCCT intensive and conventional treatment groups in the change in all biomarkers across time segments.

Conclusions—Heightened antioxidant activity (rather than diminished oxidative stress markers) is associated with lower cardiovascular disease risk in type 1 diabetes mellitus, but these biomarkers did not change over time with intensification of glycemic control.

Clinical Trial Registration—URL: https://www.clinicaltrials.gov. Unique identifiers: NCT00360815 and NCT00360893. (J Am Heart Assoc. 2018;7:e008368. DOI: 10.1161/JAHA.117.008368.)

Key Words: diabetes mellitus • F₂ iso prostaglandins • free radical • paraoxonase

The DCCT (Diabetes Control and Complications Trial) and its observational follow-up study, the EDIC (Epidemiology of Diabetes Interventions and Complications), showed that intensive therapy to lower glycated hemoglobin (HbA1c) to near-normal concentrations reduces the risk of any cardiovascular disease (CVD) event in patients with type 1 diabetes mellitus (T1DM).¹–³ Despite the landmark study’s conclusion that intensive therapy has long-term beneficial effects on the risk of CVD,⁴ the precise mechanisms by which cardiovascular complications of diabetes mellitus occur are still not well understood. Early studies have focused on the changes in lipoprotein levels as biomarkers for diabetes mellitus complications; specifically, oxidative modification of low-density lipoprotein is believed to play a causative role in the development of atherosclerotic disease as well as microvascular complications of T1DM.⁵–⁹ Although lipoprotein markers have proved to be important for assessing cardiovascular risk, they do not completely account for the excess cardiovascular risk in diabetes mellitus.
Clinical Perspective

What Is New?

- We observed that heightened antioxidant activity (as measured by paraoxonase-1 activity), rather than diminished markers of oxidative stress, is associated with lower cardiovascular disease risk in patients with type 1 diabetes mellitus.
- We also observed that none of the biomarkers improved over time with intensification of glycemic control.

What Are the Clinical Implications?

- Hyperglycemia has been associated with increased oxidative stress, which has long been implicated in the increased risk for the development of macrovascular and microvascular complications in patients with type 1 diabetes mellitus.
- These findings challenged past reports and suggest that intrinsic counter-regulatory mechanisms such as paraoxonase-1 activity may play an important role in cardioprotection in patients with type 1 diabetes mellitus.

Hyperglycemia leading to increased oxidative stress has been implicated as a key pathophysiological factor of macrovascular complications in T1DM. Through the use of specific measures of oxidative stress in the setting of known CVD-reducing therapies such as 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors, strong correlations between distinct oxidative pathways and atherosclerotic disease have been established in the general population. Leukocyte-derived oxidant production is triggered by granulocyte peroxidases such as myeloperoxidase (MPO). Furthermore, reactive species produced by activated neutrophils and other sources can lead to lipid peroxidation of the cellular membrane and the production of prostaglandin-like compounds such as F2-isoprostane (F2IP) and its major metabolite, 2,3 dinor-8 isoprostaglandin F2α (2,3-dinor-iPF2α-III). Conversely, important high-density lipoprotein components such as paraoxonases (PON) are antioxidants that can prevent the accumulation of lipid peroxides.

Finally, the widely distributed antioxidant enzyme, PON1, is closely associated with HDL. Low levels of PON1 have consistently been associated with susceptibility to coronary heart disease in various case-control studies. Additionally, mouse models of PON1 overexpression have suggested the ability of PON to inhibit development of atherosclerosis in metabolic syndrome. Because these associations have not been fully explored in T1DM, we assess the link between oxidative pathways and the risk for CVD as well as its relationship to chronic glycemic control in a longitudinal case-cohort substudy of the DCCT/EDIC patient population.

Methods

The data, analytical methods, and study materials will be/have been made available to other researchers through the National Institute of Diabetes and Digestive and Kidney Disease Data Repository for purposes of reproducing the results or replicating the procedure. These include the multiply imputed data sets that were constructed to account for missing data and the computer programs used to generate the tables herein.

Study Population

The DCCT was a randomized controlled trial of 1441 patients who were between the ages of 13 and 39 years. Approximately half the subjects were enrolled in a primary prevention cohort with no pre-existing microvascular complications and 1 to 5 years’ duration of T1DM, and half to a secondary intervention cohort with mild pre-existing retinopathy, possible microalbuminuria, and 1 to 15 years’ duration preceding study entry. At baseline, patients with a history of CVD, hypertension (blood pressure of 140/90 mm Hg or more), or hypercholesterolemia (fasting serum cholesterol 3 SDs above age- and sex-specific means) were excluded. Participants were assigned to either intensive therapy aimed at achieving chronic glycemic levels as close to the nondiabetic range as safely possible, or to conventional therapy that aimed to avoid symptomatic hyperglycemia and hypoglycemia. The cohort was followed for an average of 6.5 years. Following DCCT, 97% of the surviving original cohort agreed to join the observational follow-up study, the EDIC. The DCCT and EDIC studies were approved by the institutional review boards of all participating institutions, and each participant provided written informed consent.

Case-Cohort Design

A case-cohort design was used to assess the association of oxidative biomarkers of interest with CVD cases who had experienced an incident CVD event during the DCCT or during the first 16 years of follow-up in EDIC. CVD events were defined as fatal or nonfatal myocardial infarction or stroke and death judged to be attributed to CVD, subclinical myocardial infarction present on an annual ECG, angina confirmed by ischemic changes on exercise tolerance testing, or by clinically significant obstruction on coronary angiography or coronary bypass.

A sample of 125 cases with a sample of 250 control subjects would provide 85% power to detect a hazard ratio (approximately equal to an odds ratio) of 1.39 per SD difference using a score test in the stratified Cox regression model at the 0.05 level 2-sided.

At the time that the sample was selected, a total of 127 subjects had experienced at least 1 CVD event (cases). A
subcohort of 350 subjects was randomly selected from the DCCT cohort of 1441 subjects. No specimens were available for 1 patient, who was deleted. This subcohort included 29 CVD cases, leaving 320 controls without past CVD. To this, we added the remaining 98 then-known CVD cases, yielding a study sample of 447 subjects (127 CVD cases + 320 controls).

**Biospecimens**

Patient blood and urine samples were collected as part of the DCCT/EDIC study. Oxidative stress markers were measured on stored biosamples from 4 time points: DCCT baseline, DCCT year 1, DCCT closeout (mean 6.5 years of treatment), and EDIC years 1 to 2.

**Laboratory Analysis**

HbA1c was measured quarterly during the DCCT and annually during EDIC using a high-precision, high-performance liquid chromatography assay with long-term control subjects to monitor assay stability as described. During DCCT, fasting lipid and serum creatinine levels and other risk factors for CVD were measured annually. During EDIC, fasting lipid levels and renal function were measured in alternate years.

Serum paraoxonase activity was determined using spectrophotometry in a 96-well plate format (Spectramax 384 Plus; Molecular Devices, Sunnyvale, CA) as previously described. Rate of generation of paranitrophenol was determined at 405 nm in 40-fold diluted serum (final) in reaction mixtures composed of 1.5 mmol/L of paraoxon (Sigma-Aldrich, St. Louis, MO), 10 mmol/L of Tris hydrochloride, pH 8, 1 mol/L of sodium chloride, and 2 mmol/L of calcium chloride at 24°C. An extinction coefficient (at 405 nm) of 17 000 mol/L/cm2 was used for calculating units of paraoxonase activity, which is expressed as nmol/L of paranitrophenol produced per minute per milliliter of serum. The intra-assay and interassay coefficients of variance (CVs) for the high-throughput PON1 activity assay were 1.9% and 3.3%, respectively, on 30 replicates performed on 15 different days. Plasma concentrations of MPO were determined by MPO assay on a Siemens Dimension XPand analyzer for EDTA plasma, with intra-assay CV of 2.2% and interassay CV of 3.5%. Quantitation of urine metabolites F2-IP and 2,3-dinor-iPF2α-III were measured by stable isotope dilution liquid chromatography/tandem mass spectrometry analysis as previously described. The interday assay CV reported for the 2 assays averaged 10%, whereas the interday assay CV averaged 10.6%.

**Statistical Analysis**

Owing to depletion of saved specimens from past DCCT/EDIC ancillary studies, 261 of the 1759 (15%) expected samples were missing, 255 owing to depletion and only 6 owing to loss to follow-up. To address these missing data, multiple imputation was used to provide 10 estimates of each missing value and yielding 10 complete data sets. A given analysis was then repeated using each of the 10 complete data sets, and the results were averaged using the methods of Rubin and Schenker. The resulting confidence limits and P values accounted for the overall extent of the missing original data.

A survey sampling analysis using weights inversely proportional to the sampling probability was used to describe the characteristics of the weighted subsample in comparison to the full cohort. Mean values of the biomarkers over time with tests of treatment group differences were obtained using survey sampling regression models.

Separate analyses of the association of each natural log-transformed biomarker with the risk of an initial CVD event were conducted using Barlow’s modified Cox proportional hazards model for application to a case-cohort design that used the survey weights equal to the inverse of the subcohort sampling probabilities. Cox proportional hazards models within each group were stratified by baseline cohort and the models in the combined groups were also stratified by treatment group. All associations are presented as the percent change in CVD risk per 10% increase in the biomarker, with and without adjustment for HbA1c as a time-dependent covariate. The biomarker entered the model as a time-dependent covariate with the biomarker value at each of the 4 measurement times. The time-dependent HbA1c covariate value used the eligibility screening value at baseline, the mean over the first year at year 1, the mean up to closeout at closeout, and the mean over DCCT and EDIC up to EDIC year 1 or 2. Additional models tested group by biomarker interaction effects.

The Appendix S1 provides a technical description of the statistical methods and the software that were used.

**Results**

The weighted estimates of patient characteristics in the total case-cohort sample (N=447) were similar to the aggregate estimates in the original complete DCCT cohort (N=1441), and the validity of the inverse probability weighted analyses was verified (Table 1).

Table 2 describes the mean values of the biomarkers over the selected study period, with tests of significant differences within and across treatment groups and points in time. A significant difference (P<0.05) between baseline and DCCT year 1 was observed within both the intensive and conventional treatment groups (separately) for 2,3-dinor-iPF2α-III and for F2-IP, but with no significant difference between groups. Changes between DCCT closeout and EDIC year 1 to 2 were not significant within either treatment group and did not differ between groups.
Table 1. Baseline Characteristics of the DCCT/EDIC Case-Control and Total Cohorts

<table>
<thead>
<tr>
<th></th>
<th>Weighted Case-Control Cohort</th>
<th>Total DCCT Cohort</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>447</td>
<td>1441</td>
</tr>
<tr>
<td>Age, y</td>
<td>26.7 ± 7.3</td>
<td>26.8 ± 7.1</td>
</tr>
<tr>
<td>Female, %</td>
<td>46</td>
<td>47</td>
</tr>
<tr>
<td>Primary prevention cohort, %</td>
<td>51</td>
<td>50</td>
</tr>
<tr>
<td>Intensive treatment group, %</td>
<td>49</td>
<td>49</td>
</tr>
<tr>
<td>Diabetes mellitus duration, mo</td>
<td>69.2 ± 50.9</td>
<td>67.6 ± 49.9</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>23.5 ± 2.7</td>
<td>23.5 ± 2.8</td>
</tr>
<tr>
<td>HbA1c, %</td>
<td>8.8 ± 1.5</td>
<td>8.9 ± 1.6</td>
</tr>
<tr>
<td>HbA1c, mmol/mol</td>
<td>73.2 ± 16.4</td>
<td>74.0 ± 17.5</td>
</tr>
</tbody>
</table>

Unless otherwise indicated, data are means ± SD or % estimated using the survey sample weights based on the inverse sampling probabilities within strata defined by primary vs secondary cohort, intensive vs conventional treatment, and CVD case vs control. CVD indicates cardiovascular disease; DCCT, Diabetes Control and Complications Trial; EDIC, Epidemiology of Diabetes Interventions and Complications; HbA1c, glycated hemoglobin.

Primary prevention cohort and secondary intervention cohort are based on the original DCCT study design, see Methods.

Discussion

Strong evidence has tied oxidative stress to the development of CVD in the general population. In this study, we utilize biomarkers that are distinct elements of the oxidative process. There are several novel findings in this study. First, we observed that none of our oxidative stress biomarkers changed over time in either treatment group and did not differ significantly between groups during the course of the study. Second, increases in all but MPO were associated with lower risk of CVD with and without adjustment for HbA1c as a time dependent covariate (marginally for F2IP after adjustment). These results suggest that despite their associations with lower CVD risk, biomarkers of oxidative stress are largely unaffected by intensification of glycemic control in this prospectively randomized study of patients with T1DM. These findings are surprising given previous DCCT/EDIC studies showing increased oxidized low-density lipoprotein and advanced glycation end products as predictors of increased coronary artery calcification or carotid artery intimamedial thickness.5,7,8 The discrepancy may suggest different mechanisms by which modified low-density lipoprotein forms contribute to disease beyond those reflected by our panel of markers. Furthermore, carotid intimamedial thickness and coronary artery calcification are surrogate measures of atherosclerosis and do not directly indicate clinical events. For example, carotid intimamedial thickness is not recommended for routine measurement in clinical practice for risk assessment for a first atherosclerotic CVD event by recent American College of Cardiology/American Heart Association guidelines,32 whereas coronary artery calcification scores are generally weighted upward for greater calcium density. However, more-recent data have suggested that increased plaque calcium density may be inversely correlated with CVD risk.33

Similar to the oxidative marker findings, we observed an association between higher antioxidant PON activity and lower CVD risk in this cohort, especially in the conventional treatment group, both unadjusted and adjusted for HbA1c. An additional analysis (not shown) adjusted for high-density lipoprotein levels and smoking status at the time of each biomarker measurement provided concordant results.

Diminished PON activity has been linked with T1DM in several studies.34–38 Paraoxonases constitute a family of calcium-dependent esterases with 3 isoforms: PON1, PON2, and PON3, with PON1 being the primary form found in serum.39 Whereas all 3 isoforms exhibit arylesterase and paraoxonase activities, the native enzymatic activity of PON is thought to be as a lactonase that may modify several prodrugs.30 Studies have shown that both the more-abundant PON1, as well as PON3, bind to the high-density lipoprotein particle and circulate.40 These pleiotropic enzymes are genetically highly conserved across species, and their diverse roles include protection against lipid peroxidation and oxidative stress, modulation against endoplasmic reticulum stress, and regulation of cell proliferation and apoptosis.41 Diminished serum PON paraoxonase and arylesterase activities...
Table 2. Mean Values of Biomarkers Over the Study Period (DCCT Baseline, DCCT year 1, Closeout DCCT, and EDIC Year 1–2)

<table>
<thead>
<tr>
<th>Biomarker</th>
<th>DCCT Baseline</th>
<th>DCCT Year 1</th>
<th>Difference</th>
<th>Closeout DCCT</th>
<th>EDIC Year 1 to 2</th>
<th>Difference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MPO (pmol/L)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intensive</td>
<td>405.1±21.6</td>
<td>359.4±19.7</td>
<td>−45.7 (−97.2, 5.9)</td>
<td>383.15±18.5</td>
<td>351.0±15.8</td>
<td>−32.13 (−74.5, 10.2)</td>
</tr>
<tr>
<td>Conventional</td>
<td>359.97±18.0</td>
<td>374.8±20.7</td>
<td>14.8 (−31.8, 61.4)</td>
<td>368.4±15.8</td>
<td>390.4±21.8</td>
<td>22.0 (−20.0, 64.1)</td>
</tr>
<tr>
<td>Intensive vs conventional</td>
<td>−60.5 (−131.1, 10.2)</td>
<td>0.0929</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>PON activity, μmol/min/mL</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intensive</td>
<td>912.2±47.1</td>
<td>895.1±44.6</td>
<td>−17.1 (−55.7, 21.6)</td>
<td>884.3±47.7</td>
<td>884.9±49.0</td>
<td>0.64 (−57.8, 59.1)</td>
</tr>
<tr>
<td>Conventional</td>
<td>771.3±40.9</td>
<td>765.4±39.4</td>
<td>−5.9 (−38.7, 26.9)</td>
<td>747.1±40.4</td>
<td>754.2±51.1</td>
<td>7.03 (−66.4, 80.5)</td>
</tr>
<tr>
<td>Intensive vs conventional</td>
<td>−11.14 (−63.0, 40.7)</td>
<td>0.6716</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>F2α-isoprostane (pg/mg Cr)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intensive</td>
<td>1.16±0.05</td>
<td>1.31±0.06</td>
<td>0.15 (0.04, 0.26)</td>
<td>1.49±0.084</td>
<td>1.46±0.062</td>
<td>−0.02 (−0.19, 0.14)</td>
</tr>
<tr>
<td>Conventional</td>
<td>1.12±0.054</td>
<td>1.31±0.07</td>
<td>0.19 (0.07, 0.31)</td>
<td>1.27±0.063</td>
<td>1.30±0.063</td>
<td>0.04 (−0.09, 0.17)</td>
</tr>
<tr>
<td>Intensive vs conventional</td>
<td>−0.04 (−0.19, 0.12)</td>
<td>0.6191</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>2,3-dinor-iPF2α-III (pg/mg Cr)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intensive</td>
<td>9.86±0.51</td>
<td>12.7±0.94</td>
<td>2.84 (0.85, 4.83)</td>
<td>13.7±0.99</td>
<td>13.22±0.90</td>
<td>−0.46 (−2.57, 1.65)</td>
</tr>
<tr>
<td>Conventional</td>
<td>9.93±0.46</td>
<td>11.44±0.76</td>
<td>1.51 (0.1, 2.93)</td>
<td>11.51±0.61</td>
<td>11.9±0.713</td>
<td>0.37 (−0.87, 1.61)</td>
</tr>
<tr>
<td>Intensive vs conventional</td>
<td>1.33 (−1.4, 4.05)</td>
<td>0.3267</td>
<td></td>
<td></td>
<td></td>
<td>−0.83 (−3.1, 1.44)</td>
</tr>
</tbody>
</table>

Unadjusted means are presented in the table with standard errors. Differences within each treatment group for the time periods (baseline to DCCT year 1, and DCCT closeout to EDIC year 1–2) with 95% confidence intervals are shown. 2,3-dinor-iPF2α-III indicates 2,3 dinor-8 iso prostaglandin F2α; DCCT, Diabetes Control and Complications Trial; EDIC, Epidemiology of Diabetes Interventions and Complications; MPO, myeloperoxidase; PON, paraoxonase.
have been directly associated with increased circulating levels of structurally defined specific oxidized fatty acids and may provide incremental prognostic value in stable cardiac patients, even among those with no significant coronary artery stenosis by angiography who might otherwise be dismissed as low risk. Meanwhile, serum PON activity levels strongly tracked with genetic polymorphisms linked to PON1 genotype (especially Q192R), thus confirming the contribution of PON1 in these esterase activities. It has been reported that glycation and glycoxidation of PON1 substantially reduces the activity of PON1 in these esterase activities. Furthermore, our findings did not support the notion that intensification of glycemic control confers any improvement in F2IP and 2,3-dinor-iPF2-III levels over time. Whereas in T1DM, insulin action may contribute to enhancement in antioxidant effects and help explain the stability of the observed oxidative markers over time, we do not find it likely that this actually plays a large role in our study. In a previous assessment of risk factors for CVD and major atherosclerotic cardiovascular events, daily insulin dose did not emerge in the final model for CVD or major atherosclerotic cardiovascular events. Insulin dose alone did have a nominally significant association with risk of any CVD, but was not significant in a model with other factors.

In contrast to the F2IP formed as a product of numerous oxidative mechanisms, MPO serves as a catalytic source of 2,3-dinor-iPF2-III indicates 2,3-dinor-oxygenated prostaglandin F2α, CI, confidence interval; CVD, cardiovascular disease; F2IP, F2-isoprostane; HbA1c, glycated hemoglobin; MPO, myeloperoxidase; PON, paraoxonase.

<table>
<thead>
<tr>
<th>Biomarker</th>
<th>Intervention</th>
<th>% Change in Risk for a 10% Higher Value (95% CI)</th>
<th>P Value</th>
<th>Model Chi Square</th>
<th>% Change in Risk for a 10% Higher Value (95% CI)</th>
<th>P Value</th>
<th>Model Chi Square</th>
</tr>
</thead>
<tbody>
<tr>
<td>MPO</td>
<td>Intensive</td>
<td>4.2 (–2.3, 11.1)</td>
<td>0.21</td>
<td>2.2</td>
<td>4.0 (–2.3, 10.8)</td>
<td>0.22</td>
<td>6.5</td>
</tr>
<tr>
<td></td>
<td>Conventional</td>
<td>2.2 (–2.2, 6.9)</td>
<td>0.33</td>
<td>1.3</td>
<td>2.9 (–1.8, 7.8)</td>
<td>0.23</td>
<td>9.72</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>2.9 (–0.8, 6.8)</td>
<td>0.12</td>
<td>3.08</td>
<td>3.3 (–0.4, 7.2)</td>
<td>0.083</td>
<td>15.97</td>
</tr>
<tr>
<td>PON activity</td>
<td>Intensive</td>
<td>–4.8 (–9.3, –0.1)</td>
<td>0.046</td>
<td>5.9</td>
<td>–4.3 (–8.9, 0.6)</td>
<td>0.09</td>
<td>8.8</td>
</tr>
<tr>
<td></td>
<td>Conventional</td>
<td>–4.9 (–8.1, –1.6)</td>
<td>0.0036</td>
<td>10.3</td>
<td>–4.6 (–8.1, –1)</td>
<td>0.0125</td>
<td>16.9</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>–4.9 (–7.4, –2.2)</td>
<td>0.0004</td>
<td>16.1</td>
<td>–4.5 (–7.3, –1.6)</td>
<td>0.0026</td>
<td>25.5</td>
</tr>
<tr>
<td>F2IP</td>
<td>Intensive</td>
<td>–10.2 (–19.8, 0.6)</td>
<td>0.06</td>
<td>4.9</td>
<td>–9.4 (–19.4, 1.8)</td>
<td>0.097</td>
<td>8.54</td>
</tr>
<tr>
<td></td>
<td>Conventional</td>
<td>–6.6 (–15.6, 3.3)</td>
<td>0.18</td>
<td>2.9</td>
<td>–6.1 (–15.2, 4.0)</td>
<td>0.23</td>
<td>10.2</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>–8.1 (–15, –0.7)</td>
<td>0.034</td>
<td>7.4</td>
<td>–7.5 (–14.6, 0.2)</td>
<td>0.057</td>
<td>18.4</td>
</tr>
<tr>
<td>2,3-dinor-iPF2-III</td>
<td>Intensive</td>
<td>–6.1 (–11.0, –0.8)</td>
<td>0.02</td>
<td>5.4</td>
<td>–6.4 (–11.7, –0.8)</td>
<td>0.026</td>
<td>10.5</td>
</tr>
<tr>
<td></td>
<td>Conventional</td>
<td>–4.6 (–10.0, 1.1)</td>
<td>0.11</td>
<td>4.1</td>
<td>–4.5 (–9.9, 1.2)</td>
<td>0.12</td>
<td>11.6</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>–5.2 (–9, –1.4)</td>
<td>0.0086</td>
<td>9.1</td>
<td>–5.3 (–9.1, –1.4)</td>
<td>0.0092</td>
<td>21.6</td>
</tr>
</tbody>
</table>

2,3-dinor-iPF2-III indicates 2,3-dinor-oxygenated prostaglandin F2α, CI, confidence interval; CVD, cardiovascular disease; F2IP, F2-isoprostane; HbA1c, glycated hemoglobin; MPO, myeloperoxidase; PON, paraoxonase.

* All analyses used the log (biomarker). To assess the percent change in risk for a higher 10% value in the biomarker, the formula used is: 100 × (1.1|^p−1|).
CVD risk in the DCCT/EDIC cohort, even in the unadjusted model, is therefore unexpected.

Our findings in this study differ compared with previous studies and challenge the common belief of elevated oxidative markers as harmful stressors. However, recent evidence is evolving the interpretation of these markers. Prospective analysis of urinary F2IP and its metabolites has also found an inverse association with the risk of developing type 2 diabetes mellitus, a group known to be highly susceptible to CVD. As previously mentioned, a majority of evidence linking F2IP to CVD has been cross-sectional. Thus, the contrasting finding between prospective and cross-sectional elevation of F2IP suggests dynamic effects over time that may not be captured through cross-sectional studies. These observations suggest a need for re-evaluation of how we interpret systemic oxidative markers. Citing a recent hypothesis that CVD and diabetes mellitus, in part, may be mediated by the failure to generate sufficient reactive oxygen species, the inverse relationship between F2IP and CVD risk in our study can be explained by measurement of F2IP over time as a marker of systemic metabolic compensation. These oxidative marker findings appear to contrast with the observation that increased antioxidant PON activity is associated with lower CVD risk, but they may, in fact, represent the balancing act of disease modulation. However, only additional mechanistic studies will help expand our understanding. Overall, our findings present a novel view of biomarker associations in the T1DM population with a dominant pattern of higher oxidative stress marker along with increased antioxidant activity associated with diminished risk of CVD.

Conclusions

In the DCCT/EDIC study, heightened antioxidant activity (as measured by PON activity), rather than diminished markers of oxidative stress, is associated with lower CVD risk in T1DM. None of the biomarkers improved over time with intensification of glycemic control.

Appendix

The DCCT/EDIC Research Group

The members of the DCCT/EDIC Research Group at the time of this publication follow:

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Dr Lachin and Ms McGee had full access to all of the data in the study and both take responsibility for the integrity of the data and the accuracy of the data analysis. Drs Tang and Hazen and Mr Li drafted the article. Drs Hoogwerf, Lachin, and Ms McGee helped revise the article. D.M. Nathan is the editor for DCCT/EDIC publications. Industry Support: Industry contributors have had no role in the DCCT/EDIC study, but have provided free or discounted supplies or equipment to support participants’ adherence to the study: Abbott Diabetes Care (Alameda, CA), Animas (Westchester, PA), Bayer Diabetes Care (North America Headquarters, Tarrytown, NY), Becton Dickinson (Franklin Lakes, NJ), Eli Lilly (Indianapolis, IN), Extend Nutrition (St. Louis, MO), Insulet Corporation (Bedford, MA), Lifescan (Milpitas, CA), Medtronic Diabetes (Minneapolis, MN), Nipro Home Diagnostics (Ft. Lauderdale, FL), Nova Diabetes Care (Billerica, MA), Omron (Shelton, CT), Perrigo Diabetes Care (Allegan, MI), Roche Diabetes Care (Indianapolis, IN), and Sanofi-Aventis (Bridgewater NJ).

Sources of Funding

The DCCT/EDIC has been supported by cooperative agreement grants (1982–1993, 2012–2017), and contracts (1982–2012) with the Division of Diabetes Endocrinology and Metabolic Diseases of the National Institute of Diabetes and Digestive and Kidney Disease (current grant numbers U01 DK094176 and U01 DK094157), and through support by the National Eye Institute, the National Institute of Neurologic Disorders and Stroke, the General Clinical Research Centers Program (1993–2007), and Clinical Translational Science Center Program (2006–present), Bethesda, MD. The research in this ancillary study to the DCCT/EDIC was supported by National Institutes of Health grants P01HL076491, R01HL128300, and 3R01DK080732-01A1S1.

Disclosures

Dr Hazen is named as inventor on pending and issued patents held by the Cleveland Clinic relating to cardiovascular diagnostics and therapeutics. Dr Hazen is a paid consultant for P&G. Dr Hazen has received research funds from P&G, Pfizer Inc., and Roche Diagnostics. Dr Hazen has received royalty payments for inventions or discoveries related to cardiovascular diagnostics or therapeutics from Cleveland HeartLab, P&G, Siemens, Esperion, and Frantz Biomarkers, LLC. The remaining authors have no disclosures to report.
References


SUPPLEMENTAL MATERIAL
Supplemental Material:
EDIC Subset Case-cohort Analyses

Oxidative Stress and Cardiovascular Risk in Type 1 Diabetes Mellitus: Insights from the DCCT/EDIC Study


March 6, 2018

1 Case-Cohort Sampling

This study was designed to conduct 3 separate sub-studies of the association of biomarkers with the incidence of progression of retinopathy, progression of nephropathy, and of cardiovascular disease. For each outcome, the analysis with 125 cases and 250 controls would provide high power to detect an important association. The current paper and this supplemental material applies to the analysis of associations with CVD alone. At the time that the sampling was conducted there were a total of 127 subjects who had experienced a CVD event (the cases). A random sample of the complete cohort (the sub-cohort) of 350 subjects was selected. One the subjects did not have any remaining specimens and was not included in the study. Among the remaining 349 subjects, 29 were CVD cases leaving 320 CVD controls. Thus, the study sample comprised the 127 CVD cases and the 320 controls, 447 total. To determine the inverse sampling weights, the sampling fractions were employed within the 4 principal design strata defined by membership in the primary prevention versus secondary intervention cohort and assignment to the intensive versus conventional therapy.

2 Survey Sampling Analyses

Since a case-cohort design starts with a random sample of the full cohort, augmented by additional cases in the full cohort, the analysis must be conducted using weights inversely proportional to the sampling probabilities for cases and controls. These are called Hurwitz-Thompson estimators. Further, the expressions for the variance (standard error) of the estimators differ from those from simple random sampling. Thus, the analysis was conducted using sample survey methods that employ the inverse sampling weights. Doing so, the analysis provides estimates of the analysis results that would be expected had the full cohort been available for analysis.

3 Missing Data and Multiple Imputation

Unfortunately, there was a modest fraction of missing data. For the biomarker PON, the following table shows the numbers expected and missing at each point in time and the reason, either due to depletion or...
The patterns missing for other biomarkers measured in plasma or in urine were similar. All total, about 15% of the expected data is missing, allowing for CVD outcomes after which missing measures have no effect on the analysis. Virtually all of these are missing owing to depletion of material in preceding studies. Of these, the most troubling was the 22% missing the baseline value.

Accordingly the statistical technique of multiple imputation was employed to address the potential impact of these missing data (Rubin, 1987; van Buuren, 2012). Briefly, an imputation model is developed based on regression models for each variable as a function of the others, and the model is then applied to each missing value of each variable to impute a probable value for that instance based on the conditional expectation of the variable value plus a random error, the latter to preserve the variance and correlation structure of the data. For these data the imputation was conducted using the MICE procedure (Ragunathan et al. 2001) implemented in the R-function mice (White et al., 2011). The varables used included included age, sex, duration of diabetes, DCCT primary versus secondary cohort, BMI, smoking, HDL, LDL, MBP, log(AER), and log(mean updated HbA1c) in all imputations (using the option “include”in function mice), and the CVD, retinopathy and renal status, as well as the four oxidative stress markers (all four on the log scale) based on their association with the variable being imputed (using option “minpuc” set to 0.2 in function mice).

Ten complete data sets (with no missing values) were generated and all analyses were then conducted separately using each data set. The results of the 10 analyses were then averaged using the method of Rubin and Schenker (1986).

4 Baseline Characteristics (Table 1)

Table 1 of the paper shows the weighted estimates (mean, proportion) of patient characteristics taking into account the inverse sampling probability weights.

<table>
<thead>
<tr>
<th>Treatment Group</th>
<th>Cohort</th>
<th>CVD case/control</th>
<th>Weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intensive</td>
<td>Primary</td>
<td>Control</td>
<td>328/55</td>
</tr>
<tr>
<td></td>
<td>Secondary</td>
<td>Control</td>
<td>331/119</td>
</tr>
<tr>
<td>Conventional</td>
<td>Primary</td>
<td>Control</td>
<td>345/55</td>
</tr>
<tr>
<td></td>
<td>Secondary</td>
<td>Control</td>
<td>310/91</td>
</tr>
<tr>
<td>All</td>
<td>All</td>
<td>Case</td>
<td>127/127</td>
</tr>
</tbody>
</table>

This allows for the sampling probability of controls for each stratum defined by treatment group and cohort. Since all known CVD cases were included, the weight for all cases was 1. The sum of the weights in the first 4 strata (all controls), i.e. the sums of the numerators and denominators, equals 1314/340 and adding the 127 cases yields 1441/447.

The STATA `svy: mean` command provides an estimate of the population mean ($\hat{\mu}$) and an estimate of the variance of $\hat{\mu}$ [$\hat{V}(\hat{\mu})$] accounting for the survey design used to collect the data as captured in the above weights. Note that for a binary characteristic (e.g. female vs male) the mean is in fact a proportion. In addition, however, we also desired an estimate of the underlying standard deviation of the observations. Since the variance of the estimate is $\sigma^2/n$, where $\sigma^2$ is the simple variance, then the estimate of the population standard deviation was simply $\sigma = n\hat{V}(\hat{\mu})$ that is reported using the "estat sd" command.
5 Longitudinal Biomarker Models (Table 2)

Table 2 presents the longitudinal analysis of each biomarker as the dependent variable. Since patients entered the study with an HbA1c of about 9% and the intensive group lowered that to 7% during the first year, one question is whether there was a significant change in the biomarker from year 1 to 2 and whether this change differed between groups. Then at DCCT closeout subjects were referred to their own healthcare providers and during during the first 1-2 years the HbA1c dropped from about 9% to 8% in the conventional group and increased from about 7% to 8% in the intensive group. Thus another question was whether there was a significant change in the biomarker from year 3 to 4 and whether this change differed between groups.

This analysis was conducted using the SAS procedure SURVEYREG in which the patient was a cluster containing the measures at the 4 time points. The model then contained an effect (class) for group (intensive/conventional), time (1, 2, 3 or 4) and their interaction. The model used the weights as shown for Table 1.

The estimate function was then used to compute contrasts among the model coefficients to estimate specific effects such as the change in the biomarker from year 1 to 2 in each group (separately) and the difference between groups. The estimate statement contrast coefficients were derived using the methods described in Littell, Freund and Spector (1991).

6 Cox Proportional Hazards Models (Table 3)

Table 3 of the paper then presents prospective Cox Proportional Hazards (PH) models of the association of each biomarker with the risk of CVD. These models used the biomarker values at times 1 - 4 as a time dependent covariate. The models used the method of Barlow (1994) as implemented by Therneau and Li (1999). The models used “start-stop” notation to indicate the day (since randomization) that the biomarker value was updated. The models used an offset equal to the log of the weights. The Lin-Wei (1989) robust estimate of the covariance matrix of the model coefficients was employed to compute confidence limits and p-values.

The case-cohort design was originally described by Prentice (1986) who provided a generalization of the original PH model to conduct the analysis. Then Barlow (1994) described a simpler method of analysis that could be implemented using available software such as the SAS PROC PHREG.

Let $u_i$ be an indicator variable to denote membership in the subcohort, $= 1$ if yes, 0 otherwise; $\delta_i$ denote whether the subject had an event at time $t_i$ or is right censored at time $t_i$. Then the Barlow (1994) analysis is based on the likelihood

$$L(\beta) = \prod_{i=1}^{n_{CC}} \left[ \frac{e^{x_i^T\beta}}{(1-u_i)e^{x_i^T\beta} + \frac{1}{\alpha} \sum_{\ell \in S(t_i)} e^{x_{\ell,i}^T\beta}} \right]^{\delta_i},$$

where $n_{CC}$ is the size of the case-cohort of cases and controls, $n$ is the size of the sub-cohort, and $\alpha = n/N$ so that the contribution of the subcohort is upweighted by $N/n$, the inverse sampling weight. In EDIC the sample weights were computed separately within strata defined by treatment group (intensive versus conventional) and study cohort (primary prevention versus secondary intervention), 4 strata in total. The weights were

<table>
<thead>
<tr>
<th>Treatment Group</th>
<th>Cohort</th>
<th>Weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intensive</td>
<td>Primary</td>
<td>348/58</td>
</tr>
<tr>
<td></td>
<td>Secondary</td>
<td>363/125</td>
</tr>
<tr>
<td>Conventional</td>
<td>Primary</td>
<td>378/58</td>
</tr>
<tr>
<td></td>
<td>Secondary</td>
<td>352/108</td>
</tr>
</tbody>
</table>

Note that the weights sum to 1441/349. One subject from the original sub-cohort of 350 randomly chosen subjects did not have any specimens remaining for assay. This subject was not employed in any analyses herein, so the sub-cohort sample size enters as 349 rather than 350 as originally selected.

The original Barlow model was fit using “extra” observations in the data set using counting process structure. This can be generalized to the case where there are separate sampling fractions for cases and
controls. It is best described by example. Consider different subjects

<table>
<thead>
<tr>
<th>i</th>
<th>u_i</th>
<th>δ_i</th>
<th>start</th>
<th>stop</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>t_1</td>
</tr>
<tr>
<td>2</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>t_2</td>
</tr>
<tr>
<td>3</td>
<td>0</td>
<td>1</td>
<td>t_3</td>
<td>t_3</td>
</tr>
</tbody>
</table>

Subject 1 is a sub-cohort control at risk for the entire time to censoring (t_1). Subject 2 is a sub-cohort case that is at risk up to the event time (t_2). Subject 3 is an extra case that was not in the subcohort. That subject is at risk only at the precise event time for that case, where t^- = t - \epsilon for some small \epsilon. So all cases have a record at risk for the instant of the event, and sub-cohort cases also are at risk up to the moment prior to the event.

With time dependent covariates, subjects 1 and 2 would have records documenting the start and stop times at which the covariate process changes over time. However, for a non-sub-cohort case, there would be only one record with (start, stop) times of (t^-_3, t_3) and where the value of the time-dependent covariate is the last recorded value.

7 Software

The multiple imputations were performed in R 3.0.0. The analyses for Table 1 were conducted using the SAS PROC SURVEYMEANS and Stata using the svy: mean command. The analyses in Table were performed using the SAS PROC SURVEYREG and also using SURVEYMEANS. The analyses in Tables 3 and 4 were generated using SAS PROC PHREG. SAS version 9.2 or 9.4 was employed, and Stata version 15

8 References


Oxidative Stress and Cardiovascular Risk in Type 1 Diabetes Mellitus: Insights From the DCCT/EDIC Study

J Am Heart Assoc. 2018;7:e008368; originally published May 2, 2018;
doi: 10.1161/JAHA.117.008368
The Journal of the American Heart Association is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Online ISSN: 2047-9980

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://jaha.ahajournals.org/content/7/10/e008368