

11-14-2013

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Recommended Citation

Stegger, M., Liu, C.M., Larsen, J., Soldanova, K., Aziz, M., Contente-Cuomo, T., Petersen, A., Vandendriessche, S., Jimenez, J., Mamma, C., van Belkum, A., Salmenlinna, S., Laurent, F., Skov, R.L., Larsen, A.R., Andersen, P.S., Price, L.B. (2013). Rapid Differentiation between Livestock-Associated and Livestock-Independent Staphylococcus aureus CC398 Clades. PLoS ONE, 8(11):e79645.

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Rapid Differentiation between Livestock-Associated and Livestock-Independent *Staphylococcus aureus* CC398 Clades

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Abstract

Staphylococcus aureus clonal complex 398 (CC398) isolates cluster into two distinct phylogenetic clades based on single-nucleotide polymorphisms (SNPs) revealing a basal human clade and a more derived livestock clade. The *scn* and *tet(M)* genes are strongly associated with the human and the livestock clade, respectively, due to loss and acquisition of mobile genetic elements. We present canonical single-nucleotide polymorphism (canSNP) assays that differentiate the two major host-associated *S. aureus* CC398 clades and a duplex PCR assay for detection of *scn* and *tet(M)*. The canSNP assays correctly identified 88 *S. aureus* CC398 isolates from a reference collection into the human and livestock clades and the duplex PCR assay correctly identified *scn* and *tet(M)*. The assays were successfully applied to a geographically diverse collection of 272 human *S. aureus* CC398 isolates. The simple assays described here generate signals comparable to a whole-genome phylogeny for major clade assignment and are easily integrated into *S. aureus* CC398 surveillance programs and epidemiological studies.

Citation: Stegger M, Liu CM, Larsen J, Soldanova K, Aziz M, et al. (2013) Rapid Differentiation between Livestock-Associated and Livestock-Independent *Staphylococcus aureus* CC398 Clades. PLoS ONE 8(11): e79645. doi:10.1371/journal.pone.0079645

Editor: Michael Otto, National Institutes of Health, United States of America

Received: February 26, 2013; **Accepted:** October 4, 2013; **Published:** November 14, 2013

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Funding: This study was supported by the TGen Foundation and the National Institute of Allergy and Infectious Diseases, National Institutes of Health (project number 1R01AI101371-01A1). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: Alex van Belkum is currently employed by BioMérieux, Microbiology R&D, France, but this does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials.

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Introduction

Livestock has been considered the primary reservoir of methicillin-resistant *Staphylococcus aureus* (MRSA) clonal complex 398 (CC398); however, there is now strong evidence of a livestock-independent *S. aureus* CC398 clade circulating in humans that predates the livestock clade [1–4].

Epidemiological studies have shown that most livestock-associated MRSA CC398 (LA-MRSA CC398) strains colonize and transmit between humans to a lesser degree than other MRSA strains [5], although they are an important cause of infection in persons having direct contact with livestock [6–8]. The *scn* gene, encoding a staphylococcal complement inhibitor (SCIN) [9], and other genes in the immune evasion cluster (IEC) are likely to play important roles in evasion of the human innate immune response. IEC is largely absent from *S. aureus* CC398 isolates belonging to the livestock clade, whereas it is widespread among livestock-independent *S. aureus* CC398 isolates [3], which may, at least in part, explain the limited spread of livestock-associated *S. aureus*

CC398 isolates in humans. In addition, livestock-associated *S. aureus* CC398 isolates carry a number of resistance determinants, including the staphylococcal cassette chromosome *mec* (SCC*mec*) and the *tet(M)* gene encoding methicillin and tetracycline resistance, respectively [3].

The existence of two major host-associated *S. aureus* CC398 clades emphasizes the need for rapid molecular genotyping methods in epidemiological investigations and source tracking of *S. aureus* CC398. We describe here two assays for defining the phylogenetic origin of *S. aureus* CC398. Using these assays, we were able to determine the sources of *S. aureus* CC398 recovered from humans and to demonstrate the existence of several *scn*-positive LA-MRSA CC398 isolates that may be readapting to humans.

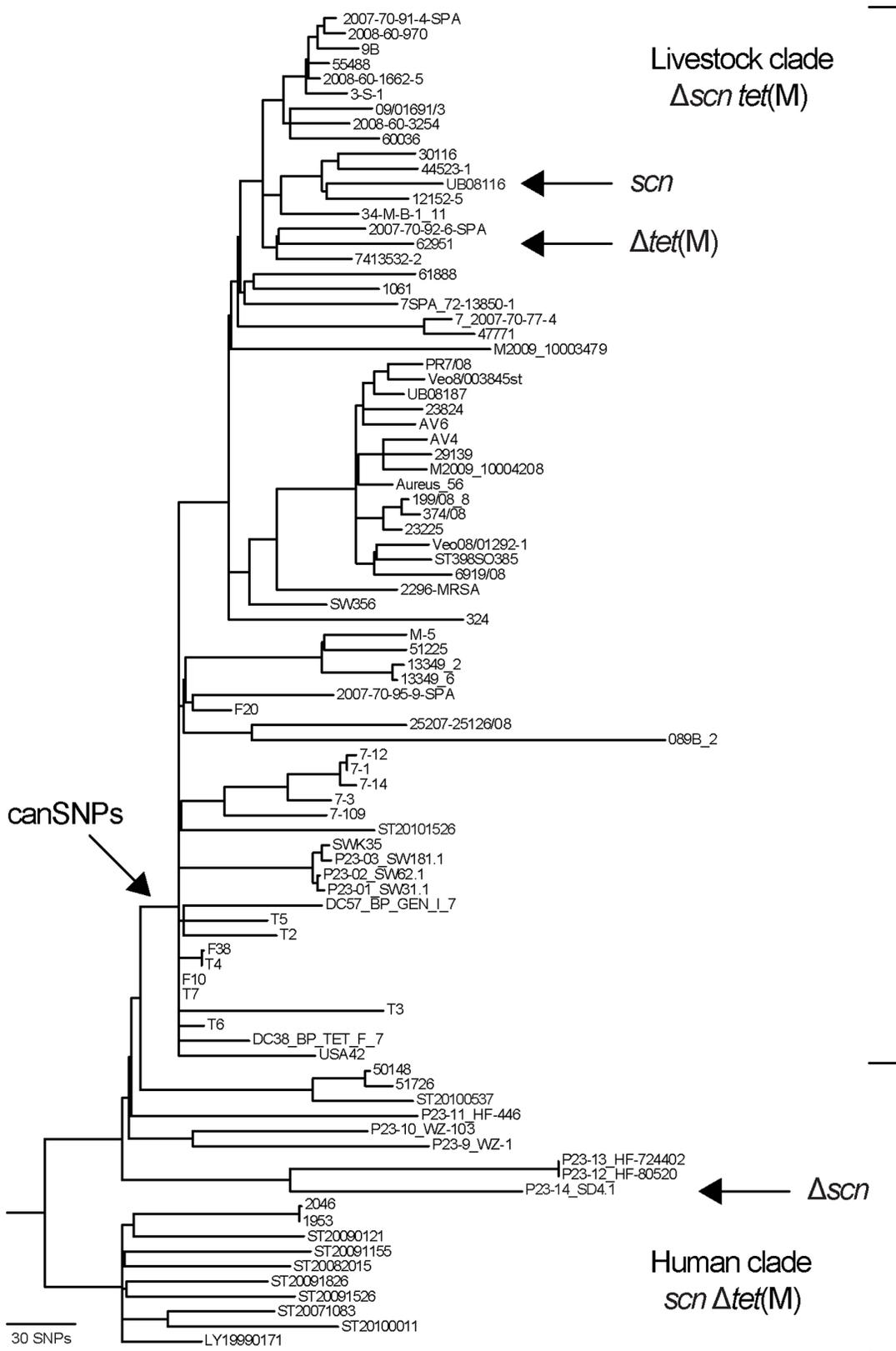


Figure 1. Maximum-parsimony tree of 89 *S. aureus* CC398 isolates based on 4,238 total SNPs, including 1,102 parsimony-informative SNPs. The bracket highlights the ancient human clade and the newly evolved livestock clade. Arrows indicate the position of the branch used to identify canSNPs, and isolates with unique *scn* and *tet(M)* patterns not consistent with the archetypal patterns are highlighted. The figure was adapted from Price *et al.* [3].
doi:10.1371/journal.pone.0079645.g001

Materials and Methods

Detection and Characterization of Single-nucleotide Polymorphisms

The phylogenetic analysis of 89 *S. aureus* CC398 core genomes identified >4,000 single-nucleotide polymorphisms (SNPs) [3]. In the present study, 13 bi-allelic, non-synonymous canonical SNPs (canSNPs) that define the two major host-associated *S. aureus* CC398 clades were identified (Table 1, Figure 1). Of these, three genetically unlinked canSNPs were selected: canSNP_748, canSNP_1002, and canSNP_3737.

S. aureus CC398 Isolates and DNA Purification

We used a reference collection of 88 *S. aureus* CC398 isolates for which phylogenetic origin and presence of the *scn* and *tet(M)* genes have been previously characterized on the basis of whole genome sequence data [3], and a collection of 272 human *S. aureus* CC398 isolates from ten countries, including Algeria (n = 2), Belgium (n = 5), Colombia (n = 1), Denmark (n = 150), Finland (n = 10), France (n = 94), India (n = 1), Italy (n = 2), Martinique (n = 2), and the Netherlands (n = 5) (Table S1). A subset of 23 isolates has been previously described in other studies [10–15].

For the 88 *S. aureus* CC398 reference isolates, DNA was purified using the DNeasy 96 Blood and Tissue Kit (QIAGEN, Valencia, CA, USA) supplemented with lysostaphin in the enzymatic lysis buffer and the Proteinase K-Buffer ATL solution. For the remaining 272 *S. aureus* CC398 isolates, DNA was obtained by incubating the bacteria in distilled water for 10 min at 95°C followed by centrifugation for 5 min at 5,000×g.

canSNP Assays

For each canSNP, 500-bp flanking regions from the chromosome of *S. aureus* CC398 reference strain SO385 (GenBank accession no. AM990992) were used to extract the corresponding

regions in the 88 whole-genome sequenced *S. aureus* CC398 isolates (Short Read Archive accession no. SRS300454–SRS300493, SRS300526–SRS300530, SRS300532, SRS300534–35, SRS300537–SRS300542, SRS300545, SRS300547, SRS300560, SRS300562–63, SRS300565, SRS300567, SRS300569, SRS300571, and SRS300580–SRS300604). The consensus sequence of each flanking region was determined using SeqMan (DNASTAR, Madison, WI, USA), and primers and fluorescently-labelled TaqMan probes were designed using Primer Express version 3.0 (Applied Biosystems, Foster City, CA, USA) (Table 2). Dual-probe real-time PCRs were performed on an ABI 7900 HT Fast Real-Time PCR System (Applied Biosystems) in 10 µL reactions, containing Platinum Quantitative PCR Super-Mix-UDG with ROX as a reference dye (Invitrogen Life Technologies, Grand Island, NY, USA), 1 µL of DNA template, 0.6 µM of each primer (Integrated DNA Technologies, San Diego, CA, USA), and 0.2 µM of each probe (Integrated DNA Technologies), with the following settings: 3 min at 50°C, 10 min at 95°C, followed by 40 cycles of 15 s at 95°C, and 1 min at 60°C. All samples were run in duplicate. Allelic discrimination files and multicomponent plots from the ABI 7900HT sequence detection system (Applied Biosystems) were visually inspected to determine the state of each canSNP.

PCR-based Detection of Adaptive Genetic Markers

The presence of the *scn* and *tet(M)* genes was investigated using a duplex PCR assay and previously published primers [16,17]; the amplicon sizes were 258 bp and 405 bp, respectively. PCRs were performed using 2 µL of template DNA. A QIAGEN Multiplex PCR Master Mix (QIAGEN) was combined with 0.2 µM of each primer in a 25 µL reaction with the following settings: 15 min at 95°C, followed by 30 cycles of 30 s at 94°C, 90 s at 52.5°C, and 90 s at 72°C, and a final extension of 10 min at 72°C. The *S. aureus* CC398 strains 50148 and 55488 were used as positive controls [3].

Results

Validation of the canSNP Assays

The canSNP assays (canSNP_748, canSNP_1002, and canSNP_3737) correctly placed 99% (87/88) of the *S. aureus* CC398 reference isolates into the human clade (n = 19) and the livestock clade (n = 68). For one isolate (F20), two assays correctly placed it in the livestock clade while one assay yielded a negative result for both states (canSNP_1002), despite the presence of conserved primer binding sites as determined by the *de novo* analysis on the whole genome sequence data.

Validation of the *scn* and *tet(M)* Duplex PCR Assay

The duplex PCR assay correctly identified the *scn* and *tet(M)* genes among the 88 *S. aureus* CC398 reference isolates. The majority (95% [18/19]) of isolates belonging to the human clade carried *scn* and lacked *tet(M)*, while a single porcine isolate (P23-14_SD4.1) lacked both genes. Conversely, most (97% [67/69]) of the isolates belonging to the livestock clade carried *tet(M)* and lacked *scn*, while one porcine isolate (62951) lacked both genes and one porcine isolate (UB08116) carried both genes. The sensitivity, specificity, and positive and negative predictive values of *scn* were 0.95, 0.99, 0.95, and 0.99, respectively, for clustering within the human clade, and those of *tet(M)* were 0.99, 1.00, 1.00, and 0.95, respectively, for clustering within the livestock clade.

Table 1. List of bi-allelic, non-synonymous canonical single-nucleotide polymorphisms (canSNPs) that define the two major host-associated *S. aureus* CC398 clades.

canSNP ^a	Genomic position ^b	Codon	
		Human clade	Livestock clade
15	9,319 (SAPIG0006)	GCC (Ala)	GTC (Val)
237	244,322 (SAPIG0223)	ATG (Met)	ATA (Ile)
476	425,594 (SAPIG0409)	CCA (Pro)	TCA (Thr)
<u>748</u>	551,946 (SAPIG053)	CCA (Pro)	TCA (Thr)
<u>1,002</u>	732,619 (SAPIG0698)	CTA (Leu)	ATA (Ile)
2,167	1,518,366 (SAPIG1434)	GCG (Ala)	GAG (Glu)
2,181	1,524,032 (SAPIG1434)	ATG (Met)	ATA (Ile)
2,761	1,934,659 (SAPIG1823)	CCA (Pro)	CTA (Leu)
3,216	2,287,341 (SAPIG2210)	ATT (Ile)	GTT (Val)
3,399	2,395,959 (SAPIG2317)	CCT (Pro)	CTT (Leu)
<u>3,737</u>	2,597,585 (SAPIG2511)	GGG (Gly)	GAG (Glu)
4,127	2,805,707 (SAPIG2701)	CGC (Arg)	TGC (Cys)
4,130	2,806,556 (SAPIG2701)	ACA (Thr)	TCA (Ser)

^acanSNPs used in the study are underlined.

^bThe genomic position was mapped to the chromosome (genes) of *S. aureus* CC398 reference strain SO385 (GenBank accession no. AM990992).

doi:10.1371/journal.pone.0079645.t001

Table 2. Primers and TaqMan probes used to identify canonical single-nucleotide polymorphisms (canSNPs) that define the two major host-associated *S. aureus* CC398 clades.

canSNP assay	Primers (5'-3')	Probes (5'-3') ^a	canSNP clade	Genomic position ^b
748	GGTACTAAGGTATATCCGTGGATTGC	6-FAM-TCTGATTTCA <u>T</u> CACCGC	Livestock	551,946 (SAPIG0537)
	ATCAGTTGCGCTAAATCTTCTATTGA	VIC-TCTGATTTCA <u>C</u> CACCGC	Human	
1,002	GAAACCAAAGGTAAAACCTAGCAAA	6-FAM-CAACAAGTGAATATAT <u>T</u>	Livestock	732,619 (SAPIG0698)
	AATTAAGCAATCGGGGTGCT	VIC-CAACAAGTGAATAT <u>T</u>	Human	
3,737	TTAYATATTTTTGGTTAACATCTTGCC	6-FAM-TTTAACTTTT <u>G</u> AGTTAGTAGCT	Livestock	2,597,585 (SAPIG2511)
	AAAATAGCTAGTGAATAATAACTGCGAGT	VIC-TTTAACTTTT <u>G</u> GTTAGTAGCT	Human	

^aTaqMan probes for the specific allele of the two canSNP states (underlined) were fluorescently labeled with either 6-FAM or VIC dye.

^bThe genomic position was mapped to the chromosome (genes) of *S. aureus* CC398 strain SO385 (GenBank accession no. AM990992).

doi:10.1371/journal.pone.0079645.t002

Application of Assays

Applying the canSNP assays (canSNP_748, canSNP_1002, and canSNP_3737) on a collection of 272 human *S. aureus* CC398 isolates produced congruent results for 97% (265/272) of the *S. aureus* CC398 isolates. For seven isolates, two of the three assays placed them in the human clade while one assay yielded a negative result for both states, including canSNP_1002 (n = 6) and canSNP_3737 (n = 1). Using a two-out-of-three rule for defining *S. aureus* CC398 isolates, the canSNP assays placed the *S. aureus* CC398 isolates into the human clade (n = 111) and the livestock clade (n = 161). All (100% [111/111]) isolates belonging to the human clade carried *scn* and lacked *tet(M)*, whereas most (96% [155/161]) of the isolates belonging to the livestock clade carried *tet(M)* and lacked *scn*. The remaining six isolates belonging to the livestock clade either carried both genes (n = 4) or lacked both genes (n = 2).

Discussion

We recently showed that the livestock-associated *S. aureus* CC398 clade evolved from the basal human clade, and that this human-to-livestock host jump was accompanied by the loss of a bacteriophage (Φ Sa3) harboring *scn* and functionally related genes that encode modulators of human innate immunity (IEC) and acquisition of a Tn916-like transposon carrying the *tet(M)* gene that confers resistance to tetracycline, which is commonly used in livestock production [3].

In the present study, we developed and validated two rapid molecular genotyping methods for defining the two major host-associated *S. aureus* CC398 clades. The three canSNP assays were developed to run optimally under identical conditions and were not affected by the DNA extraction method. Three unrelated SNP positions were included to minimize risk of false clade assignment due to the risk of nucleotide reversal or horizontal gene transfer. Nearly all of the isolates were assigned to the same clade by all three canSNP assays, and only a small subset of isolates yielded a negative result in one of the three assays. To avoid potential misclassifications by the canSNP assays, we suggest instituting a two-out-of-three rule for defining *S. aureus* CC398 isolates. As expected, most of the isolates that were assigned to the human clade by use of the canSNP assays carried *scn* and lacked *tet(M)*, while the majority of isolates that were placed in the livestock clade carried *tet(M)* and lacked *scn*.

S. aureus CC398 has been identified in humans with no apparent livestock-associated risk factors in several geographically diverse areas, including the People's Republic of China [18], Denmark

[19], France [20,21], French Guiana [22], the Caribbean [23,24], and the United States [23,25,26]. Subsequently, whole-genome sequence analysis of *S. aureus* CC398 isolates from these geographic areas demonstrated that they belong to the human clade [3,4]. By use of the assays reported here, we identified the first cases of *S. aureus* CC398 belonging to the human clade in Algeria, Belgium, Finland, India, and the Netherlands. The majority of these cases had no prior exposure to livestock [10,12,15]. These results underscore the usefulness of integrating these assays into *S. aureus* CC398 surveillance programs and epidemiological studies.

IEC is present at a high frequency among *S. aureus* clones circulating in humans but appears to have been lost during multiple independent human-to-animal host jumps by *S. aureus* belonging to CC398 [3], CC5 [27], CC97 [28], and CC8 [29]. In addition, IEC is absent from a porcine *S. aureus* CC398 isolate (P23-14_SD4.1) belonging to the human clade [3]. The independent loss of IEC in multiple *S. aureus* lineages provides strong support for the view that IEC is undergoing purifying selection in animal hosts. By contrast, the presence of IEC in a porcine *S. aureus* CC398 isolate displaying *spa* type t899 (UB08116) within the livestock clade supports that IEC has been reacquired [3]. In the present study, we also identified IEC in four human LA-MRSA CC398 isolates that were placed in the livestock clade by the canSNP assays, including three isolates displaying *spa* type t899 from Denmark and France and one isolate displaying *spa* type t034 from Denmark. Of note, a recent study provided support for the view that acquisition of IEC has facilitated animal-to-human host jumps by livestock-associated *S. aureus* CC97 isolates, leading to community spread worldwide [28]. It is therefore possible that reacquisition of IEC enables livestock-associated *S. aureus* CC398 to spread in human populations in a livestock-independent manner. We are currently monitoring for livestock-associated *S. aureus* CC398 harboring IEC in persons with and without livestock exposure to assess the risk for the emergence of a sustainable community reservoir for livestock-associated *S. aureus* CC398.

Based on the presence/absence pattern of *tet(M)*, it is tempting to speculate that the tetracycline resistance phenotype can be used as a marker for *S. aureus* CC398 isolates belonging to the livestock clade. However, all *S. aureus* CC398 isolates from Finland that were placed in the human clade by use of the canSNP assays (n = 5) were resistant to tetracycline [12]; these isolates were subsequently shown to carry the tetracycline resistance gene *tet(K)* rather than the *tet(M)* gene (unpublished data). Thus, whereas tetracycline susceptibility may have utility as a screening tool to exclude *S. aureus* CC398 isolates belonging to the livestock clade,

the tetracycline resistance phenotype seems to be less useful for exclusion of isolates belonging to the human clade.

In conclusion, the present study has underscored the usefulness of two molecular genotyping methods for defining the major host-associated *S. aureus* CC398 clades and has illustrated the power of integrating surveillance, molecular epidemiology, bioinformatics, and microbiology. Results from the two methods can be used independently for epidemiological investigations and source tracking and can be combined to screen for evolutionary signs of adaptation to new hosts and to predict public health risk. Integrating the assays into surveillance programs will aid in determining which reservoirs and bacterial factors are responsible for the increasing prevalence of *S. aureus* CC398 in the community.

References

- Davies PR, Wagstrom EA, Bender JB (2011) Lethal necrotizing pneumonia caused by an ST398 *Staphylococcus aureus* strain. *Emerg Infect Dis* 17: 1152–3.
- McCarthy AJ, van Wamel W, Vandendriessche S, Larsen J, Denis O, et al (2012) A *Staphylococcus aureus* CC398 clade associated with human-to-human transmission. *Appl Environ Microbiol* 78: 8845–8.
- Price LB, Stegger M, Hasman H, Aziz M, Larsen J, et al (2012) *Staphylococcus aureus* CC398: host adaptation and emergence of methicillin resistance in livestock. *MBio* 3: e00305–11.
- Uhlemann AC, Porcella SF, Trivedi S, Sullivan SB, Hafer C, et al (2012) Identification of a highly transmissible animal-independent *Staphylococcus aureus* ST398 clone with distinct genomic and cell adhesion properties. *MBio* 3: e00027–12.
- Wassenberg MW, Bootsma MC, Troelstra A, Kluytmans JA, Bonten MJ (2011) Transmissibility of livestock-associated methicillin-resistant *Staphylococcus aureus* (ST398) in Dutch hospitals. *Clin Microbiol Infect* 17: 316–319.
- Cuny C, Nathaus R, Layer F, Strommenger B, Altmann D, et al (2009) Nasal colonization of humans with methicillin-resistant *Staphylococcus aureus* (MRSA) CC398 with and without exposure to pigs. *PLoS One* 4: e6800.
- Graveland H, Wagenaar JA, Bergs K, Heesterbeek H, Heederik D (2011) Persistence of livestock associated MRSA CC398 in humans is dependent on intensity of animal contact. *PLoS One* 6: e16830.
- van Cleef BAGL, Monnet DL, Voss A, Krzivanek K, Allerberger F, et al (2011) Livestock-associated methicillin-resistant *Staphylococcus aureus* in humans, Europe. *Emerg Infect Dis* 17: 502–5.
- Rooijackers SHM, Ruyken M, Roos A, Daha MR, Presanis JS, et al (2005) Immune evasion by a staphylococcal complement inhibitor that acts on C3 convertases. *Nat Immunol* 6: 920–7.
- Vandendriessche S, Kadlec K, Schwarz S, Denis O (2011) Methicillin-susceptible *Staphylococcus aureus* ST398-t571 harbouring the macrolide-lincosamide-streptogramin B resistance gene *erm(T)* in Belgian hospitals. *J Antimicrob Chemother* 66: 2455–9.
- Jimenez JN, Velez LA, Mediavilla JR, Ocampo AM, Vanegas JM, et al (2011) Livestock-associated methicillin-susceptible *Staphylococcus aureus* ST398 infection in woman, Colombia. *Emerg Infect Dis* 17: 1970–1.
- Salmenlinna S, Lyytikäinen O, Vainio A, Myllyniemi AL, Raulo S, et al (2010) Human cases of methicillin-resistant *Staphylococcus aureus* CC398, Finland. *Emerg Infect Dis* 16: 1626–9.
- Mammina C, Calà C, Plano MR, Bonura C, Vella A, et al (2010) Ventilator-associated pneumonia and MRSA ST398, Italy. *Emerg Infect Dis* 16: 730–1.
- Mammina C, Bonura C, di Carlo P, Calà C, Aleo A, et al (2010) Daptomycin non-susceptible, vancomycin intermediate methicillin-resistant *Staphylococcus aureus* ST398 from a chronic leg ulcer, Italy. *Scand J Infect Dis* 42: 955–7.
- van Belkum A, Melles DC, Peeters JK, van Leeuwen WB, van Duijkeren E, et al (2008) Methicillin-resistant and -susceptible *Staphylococcus aureus* sequence type 398 in pigs and humans. *Emerg Infect Dis* 14: 479–83.
- van Wamel WJ, Rooijackers SH, Ruyken M, van Kessel KP, van Strijp JA (2006) The innate immune modulators staphylococcal complement inhibitor and

Supporting Information

Table S1 Molecular characteristics of 272 human *Staphylococcus aureus* CC398 isolates.
(XLSX)

Acknowledgments

We would like to thank Stine Frese-Madsen, Lone Ryste Hansen Kildevang and Julie Hindsberg Nielsen for excellent technical assistance.

Author Contributions

Conceived and designed the experiments: MS CML JL MA RLS PSA LBP. Performed the experiments: MS JL KS TC. Analyzed the data: MS JL KS TC PSA LBP. Contributed reagents/materials/analysis tools: AP SV JNJ CM AvB SS FL ARL. Wrote the paper: MS CML JL PSA LBP.

- chemotaxis inhibitory protein of *Staphylococcus aureus* are located on beta-hemolysin-converting bacteriophages. *J Bacteriol* 188: 1310–5.
- Warsa UC, Nonoyama M, Ida T, Okamoto R, Okubo T, et al (1996) Detection of *tet(K)* and *tet(M)* in *Staphylococcus aureus* of Asian countries by the polymerase chain reaction. *J Antibiot (Tokyo)* 49: 1127–32.
 - Yu F, Chen Z, Liu C, Zhang X, Lin X, et al (2008) Prevalence of *Staphylococcus aureus* carrying Panton-Valentine leukocidin genes among isolates from hospitalised patients in China. *Clin Microbiol Infect* 14: 381–4.
 - Lewis HC, Molbak K, Reese C, Aarestrup FM, Selchau M, et al (2008) Pigs as source of methicillin-resistant *Staphylococcus aureus* CC398 infections in humans, Denmark. *Emerg Infect Dis* 11: 1383–9.
 - Rasigade JP, Laurent F, Hubert P, Vandenesch F, Etienne J (2010) Lethal necrotizing pneumonia caused by an ST398 *Staphylococcus aureus* strain. *Emerg Infect Dis* 16: 1330.
 - van der Mee-Marquet N, Francois P, Domelier-Valentin AS, Coulomb F, Decreux C, et al (2011) Emergence of unusual bloodstream infections associated with pig-borne-like *Staphylococcus aureus* ST398 in France. *Clin Infect Dis* 52: 152–3.
 - Ruimy R, Angebault C, Djossou F, Dupont C, Epelboin L, et al (2010) Are host genetics the predominant determinant of persistent nasal *Staphylococcus aureus* carriage in humans? *J Infect Dis* 202: 924–34.
 - Bhat M, Dumortier C, Taylor BS, Miller M, Vasquez G, et al (2009) *Staphylococcus aureus* ST398, New York City and Dominican Republic. *Emerg Infect Dis* 15: 285–7.
 - Uhlemann AC, Porcella SF, Trivedi S, Sullivan SB, Hafer C, et al (2012) Molecular characterization of *Staphylococcus aureus* from outpatients in the Caribbean reveals the presence of pandemic clones. *Eur J Clin Microbiol Infect Dis* 31: 505–11.
 - Mediavilla JR, Chen L, Uhlemann AC, Hanson BM, Rosenthal M, et al (2012) Methicillin-susceptible *Staphylococcus aureus* ST398, New York and New Jersey, USA. *Emerg Infect Dis* 18: 700–2.
 - Uhlemann AC, Hafer C, A Miko B, G Sowash M, Sullivan SB, et al (2013) Emergence of sequence type 398 as a community- and healthcare-associated methicillin-susceptible *Staphylococcus aureus* in northern Manhattan. *Clin Infect Dis* 57: 700–3.
 - Lowder BV, Guinane CM, Ben Zakour NL, Weinert LA, Conway-Morris A, et al (2009) Recent human-to-poultry host jump, adaptation, and pandemic spread of *Staphylococcus aureus*. *Proc Natl Acad Sci U S A* 106: 19545–50.
 - Spoor LE, McAdam PR, Weinert LA, Rambaut A, Hasman H, et al (2013) Livestock origin for a human pandemic clone of community-associated methicillin-resistant *Staphylococcus aureus*. *mBio* 4: e00356–13.
 - Resch G, François P, Morisset D, Stojanov M, Bonetti EJ, et al (2013) Human-to-bovine jump of *Staphylococcus aureus* CC8 is associated with the loss of a β -hemolysin converting prophage and the acquisition of a new staphylococcal cassette chromosome. *PLoS One* 8: e58187.